Design and Synthesis of New Potent *C***2-Symmetric HIV-1 Protease Inhibitors. Use of L-Mannaric Acid as a Peptidomimetic Scaffold**

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A study on the use of derivatized carbohydrates as C_2 -symmetric HIV-1 protease inhibitors has been undertaken. L-Mannaric acid (**6**) was bis-*O*-benzylated at C-2 and C-5 and subsequently coupled with amino acids and amines to give C_2 -symmetric products based on C-terminal duplication. Potent HIV protease inhibitors, **28** $K_i = 0.4$ nM and **43** $K_i = 0.2$ nM, have been discovered, and two synthetic methodologies have been developed, one whereby these inhibitors can be prepared in just three chemical steps from commercially available materials. A remarkable increase in potency going from $IC_{50} = 5000$ nM (23) to $IC_{50} = 15$ nM (28) was observed upon exchanging $-COOMe$ for $-CONHMe$ in the inhibitor, resulting in the net addition of one hydrogen bond interaction between each of the two $-NH-$ groups and the HIV protease backbone (Gly 48/148). The X-ray crystal structures of **43** and of **48** have been determined (Figures 5 and 6), revealing the binding mode of these inhibitors which will aid further design.

Introduction

The human immunodeficiency virus (HIV) has been identified as the etiologic agent of acquired immunodeficiency syndrome (AIDS).1 The *pol* gene of the human immunodeficiency virus type 1 (HIV-1) encodes for the aspartic protease which mediates proteolytic processing of the *gag* and the *gag-pol* viral gene products liberating functional enzymes and structural proteins which are essential for the formation of the mature, infectious virus.2 Inactivation of the aspartic protease leads to the formation of noninfectious virions.3 As a result the HIV-1 protease has become one of the major targets for therapeutic intervention in AIDS and in HIV infection. Recently four protease inhibitors, saquinavir (**1**), ritonavir (**2**), indinavir (**3**), and nelfinavir (**4**) were approved for clinical use by the FDA (Figure 1). Despite the clinical efficacy the benefits from long-term treatment with these agents remains to be demonstrated. Selection of a drug resistant mutant virus is likely to occur after prolonged treatment reducing efficacy of therapy. The high cost of synthesis is today a barrier to the widespread use of the currently approved protease inhibitors, notably in less developed countries. It is thus important that new improved protease inhibitors accessible at low costs are developed.4

Numerous reports describing potent protease inhibitors have been disclosed.⁵ On the basis of X-ray crystal

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structures it has been shown that the HIV-1 protease exists as a *C*₂-symmetric dimer. Recognizing that both the N- and C-terminal of a substrate or inhibitor binds into identical subsites of the HIV protease and that duplication of either the N- or the C-terminal produces a *C*₂-symmetric compound has led to the design and synthesis of new inhibitors.⁵ A successful exploration of the N-terminal duplication concept was demonstrated for the (3*R*,4*R*)-diaminodiol **5** and for a related series of compounds.⁶

We have recently explored D-mannitol as a linear peptidomimetic scaffold7 and have shown that D-mannitol based *C*2-symmetric protease inhibitors of the general structure **6** have antiviral activities similar to the diaminodiol inhibitors **5** (Figure 2).8 Cyclic protease inhibitors derived from D-mannitol and L-mannitol have also been recently disclosed by us and others.⁹

Inhibitor Design. The present work describes the use of carbohydrates in the design and synthesis of structurally new *C*₂-symmetric protease inhibitors incorporating C-terminal duplication (Figure 3). From molecular modeling L-mannaric acid (**7**) was selected as a *C*2-symmetric backbone where *O*-benzylation of the hydroxyls at C-2 and C-5 of **7** and subsequent coupling with amino acids or amines gives inhibitors of the general structure **8** (Figure 3). While inhibitors based on N-terminal duplication have been extensively studied, there are fewer reports on *C*2-symmetric inhibitors based on C-terminal duplication.¹⁰ The absolute stereochemistry of the P1/P1′ substituents and of the central diol varies from one class of inhibitors to another, cf. A-759256 (**9**) and DMP 323,11 (**10**) (Figure 4). We have used computer-assisted molecular modeling to build and conformationally refine potential inhibitors in extended

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Saquinavir

Indinavir

Nelfinavir

Figure 1. FDA-approved HIV-1 protease inhibitors.

Figure 2.

 $conformations¹²$ and to match these with X-ray crystal structures.13 This analysis indicated that inhibitors derived from substituted L-mannaric acid would give a good fit with respect to (a) the spacial direction of the P1/P1′ substituents (benzyloxy), (b) the carbonyl oxygens of **8** hydrogen bonding to the structural water, and (c) the stereochemistry of the central hydroxyl groups forming hydrogen bonds to the carboxylic acid residues of Asp 25 and Asp 125. The amino acids or amines providing the P2/P2′ substituents were modeled extend-

Figure 4.

Scheme 1. Synthesis of Protected L-Mannaric Acid

ing into the lipophilic S2/S2′ pockets of the HIV-1 protease and/or hydrogen bonding to the inhibitor backbone. It is noteworthy that with the present inhibitor design strategy diverse P1 and P2 substituents in **8** are synthetically readily available making optimization of potential inhibitors feasible.

Results and Discussion

Chemistry. For the synthesis of inhibitors of the general structure **8** two synthetic routes were developed. In the first route (synthetic route 1) a suitably protected 2,5-di-*O*-benzylated L-mannaric acid was prepared and coupled to a set of amines and amino acid derivatives. In the second synthetic route developed (synthetic route 2), which is substantially shorter, 2,5-di-*O*-benzyl-Lmannaro-1,4:3,6-di-*γ*-lactone (**36**) was prepared and subsequently in one step reacted with various amines and amino acid derivatives giving bisamides **8** by nucleophilic ring opening of the bislactone **36**.

Synthetic Route 1. (Schemes 1 and 2) The monoacetonide **12** was prepared using a modified procedure as described for the corresponding D-mannitol acetonide.14 Reduction of L-mannonic *γ*-lactone (**11**) with lithium borohydride in dry MeOH gave L-mannitol which, after azeotropic removal of the boric acid and without further purification, was protected using 2,2 dimethoxypropane in acetone containing camphorsulfonic acid to give the triacetonide in 78% overall yield. Partial hydrolysis with aqueous HOAc (70%) provided the monoacetonide **12** in 64% yield after recrystallization. Subsequent selective protection of the primary hydroxyls with *tert*-butyldimethylsilyl chloride in pyridine, containing *p*-(dimethylamino)pyridine, afforded

compound **13** in 96% yield.15 Benzylation of the 2,5 diol using benzyl bromide, sodium hydride, and a catalytic amount of tetrabutylammonium iodide in THF gave **14** in 76% yield. Desilylation of **14** using tetrabutylammonium fluoride in THF delivered compound **15** in 98% yield. For the oxidation of primary hydroxyls to the corresponding dicarboxylic acid, the majority of published methods gives lactones when applied to 1,4-, 1,5-, or 1,6-diols. Attempted oxidation of compound **15** with Jones oxidation¹⁶ resulted in a complex product mixture, and attempts to oxidize the 1,6-diol using PDC and acetic acid anhydride¹⁷ produced a slightly more polar product which decomposed upon workup. The reagent system 2,2,6,6-tetramethylpiperidine 1-oxyl radical (TEMPO)-NaOCl has previously been employed for the direct conversion of alcohols to carboxylic acids and for the preparation of lactones from 1,4- and 1,5 diols.18 However with this reagent system, it is reported that oxidation of 1,6-diols to the corresponding lactone results in the formation of polymerization products.18 Notably, by using 0.05 molar equivalents of TEMPO and an excess of NaOCl at 0 °C, compound **15** was oxidized directly to the dicarboxylic acid **16** in 67% yield (Scheme 1). The successful implementation of this oxidation procedure seems to rely upon the presence of the cyclic 3,4-*O*-isopropylidene group which effectively prevents intramolecular side reactions. In the absence of the cyclic protection groups the dicarboxylic acid could not be isolated.

For introducing the amino acids and amines (Scheme 2) corresponding to the P2/P2′ substituents of the inhibitor, compound **16** was dissolved in CH_2Cl_2 -THF and condensed with the corresponding amino acid derivatives with HOBT-EDC. The condensation products, compounds **¹⁷**-**21**, were isolated in yields ranging from 34% to 73%. The 3,4-*O*-isopropylidene group was cleaved off with 4% HCl in MeOH, giving the corresponding diol products **²³**-**²⁷** in approximately 70% yield (Table 1). Compounds **²⁸**-**³¹** were prepared from **16** (Scheme 2, route A) in the same manner as that for the preparation of **²³**-**27,** but the intermediate monoacetonide was not isolated in this case. Valin methylamide, used in the preparation of **28**, was synthesized through amidation of Z-Val-OSu in dry THF followed by hydrogenolysis to furnish the methyl amide in 78% overall yield.19

In an alternative coupling procedure L-mannaric acid **16** was activated with *N*,*N*-disuccinimidyl carbonate20 to give the corresponding activated bissuccinimidyl ester **22** in 88% yield, which conveniently could be stored prior to use. This protocol avoids repeating the activation step before coupling with the appropriate amines (Scheme 2, route B). Other synthetic routes 21 examined to prepare the active ester **22** gave lower yields, and attempts to prepare the corresponding pentafluorophenyl diester were not successful.22 Coupling of **22** with three hydroxy anilines in CH_2Cl_2 followed by cleavage of the isopropylidene group with 4% HCl in MeOH delivered the diamide products **³²**-**³⁴** in 27-38% overall yield.

The cleavage step of the 3,4-*O*-isopropylidene groups in the synthesis of compounds **²³**-**³⁴** was very sensitive to the reaction conditions and other cleavage conditions examined, i.e., using TFA-H2O mixtures or PTS in dioxane-H2O gave complex product mixtures. Alternatively the hydrolysis was successfully executed with 2,3-dichloro-5,6-dicyano-1,4-benzoquinone (DDQ) in 9:1 $CH₃CN-H₂O²³$

Synthetic Route 2. (Scheme 3) Oxidation of Lmannonic *γ*-lactone (**11**) with aqueous nitric acid provided, after workup, L-mannaro-1,4:3,6-di-*γ*-lactone (**35**) in 60% yield.24 Benzylation of bislactone **35** was performed using benzyltrichloroacetimidate²⁵ with a catalytic amount of trifluoromethanesulfonic acid in dry dioxane furnishing the dibenzylated product **36** in 72% yield. Numerous other solvents besides dioxane were examined, but either yields were low or compound **35** could not be dissolved in the solvent. Benzylation procedures employing basic conditions including benzylation with benzylbromide and silveroxide were also examined, but all failed to give the desired product in reasonable yields probably attributed due to the base lability of **35**.

Nucleophilic ring opening of dilactone **36** with amines gave the bisamide derivatives **²⁸**, **³⁷**-**⁴⁸** in 22-76% yields (Scheme 3). Best yields were obtained in CH2- $Cl₂$ at reflux while moderate yields were obtained in chloroform, CH3CN, dioxane, or THF. Low yields were obtained in dimethylformamide and in ethanol. The major side product of the reaction results from *â*-elimination of **36** to give **49** (Scheme 4). The formation of **Table 1.** Structures, Yields, Methods of Preparation, and HIV-1 Protease Inhibititory Activity of L-Mannaric Diamides with 2*R*,3*R*,4*R*,5*R* Configuration

$A -$	Cmpd. no.	Yield $\%$	Method ^a	${IC}_{50}$ ${(\mu M)}^b$	\cdots K_i (nM) ^c	$A-$	Cmpd. no.	Yield $\frac{9}{96}$	Method ^a	$\frac{\mathsf{IC}_{50}}{\mathsf{(}\mu\mathsf{M}\mathsf{)}}^{\mathsf{b}}$	K_i (nM) ^c
MeO, o Ph	$\begin{array}{c} \mathbf{17}^\mathsf{d} \\ \mathbf{23} \end{array}$	72% 69%	I $\mathbf I$	$\sqrt{5}$	$^{\rm nd}$	OH	38	60%	IV	nd	5000
MeO,	18 ^d 24	70% 70%	I $\mathbf I$	ni	$^{\rm nd}$	ö $\begin{picture}(180,170) \put(0,0){\line(1,0){155}} \put(15,0){\line(1,0){155}} \put(15,0){\line(1,0){155}} \put(15,0){\line(1,0){155}} \put(15,0){\line(1,0){155}} \put(15,0){\line(1,0){155}} \put(15,0){\line(1,0){155}} \put(15,0){\line(1,0){155}} \put(15,0){\line(1,0){155}} \put(15,0){\line(1,0){155}} \put(15,0){\line(1,0){155$	39	61%	IV	nd	500- 1500
BnO	$\frac{19^{\text{d}}}{25}$	71% 71%	$\bf I$ $\bf I$	ni	nd	S	40	73%	${\rm IV}$	nd	660
HO	20 ^d 26	61% 74%	I $\bf I$	ni	$^{\rm nd}$	\sim_{N}					
$\begin{picture}(130,10) \put(0,0){\line(1,0){150}} \put(15,0){\line(1,0){150}} \put(15,0){\line(1,0){150}} \put(15,0){\line(1,0){150}} \put(15,0){\line(1,0){150}} \put(15,0){\line(1,0){150}} \put(15,0){\line(1,0){150}} \put(15,0){\line(1,0){150}} \put(15,0){\line(1,0){150}} \put(15,0){\line(1,0){150}} \put(15,0){\line(1,0){150}}$	$_{\rm 27}^{\rm 21}$	73% 73%	I $\mathbf I$	\mathbf{n}	nd	H. ő	41	31%	${\rm IV}$	nd	$2.3\,$
	${\bf 28}$ 28	88% 70%	$\rm II$ ${\rm IV}$	0.015	$0.4\,$		42	20%	${\rm IV}$	$^{\rm nd}$	$\bf 81$
N	29	29%	$\rm II$	ni	$^{\rm nd}$	OН	43	35%	${\rm IV}$	$^{\rm nd}$	0.2
\mathbf{R}^{λ}	30	30%	$\rm II$	ni	nd	OH	44	22%	IV	$_{\rm ni}$	nd
$\mathbf{M}_{\mathbf{y}}$	31	21%	$\rm II$	ni	nd	но-	45	49%	${\rm IV}$	\mathbf{ni}	nd
H_0	32	25%	\mathbf{III}	ni	$^{\rm nd}$		46	65%	IV	$^{\rm nd}$	7100
\mathbb{R}	33	32%	$\rm III$	ni	$^{\rm nd}$	$\sum_{\mathbf{r}} \mathbf{h}_{\mathbf{r},\mathbf{r}}$	47	59%	${\rm IV}$	$^{\rm{nd}}$	2000
H、 , HO	34	38%	$\rm III$	\mathbf{ni}	nd	$\sum_{i=1}^{n}$ $\frac{1}{\mu}$	48	46%	IV	nd	0.9
$\frac{1}{\sqrt{\frac{1}{n}}}\sum_{i=1}^{n}$ \mathbf{h}^{λ}	37	76%	${\rm IV}$	nd	140						

^a Method I: see Scheme 2, route A. Method II: see Scheme 2, route A (overall yields from **16**). Method III: see Scheme 2, route B (yields from **22**). Method IV: see Scheme 3 (amine opening of bislactone 36). b ni = no inhibition at 10 μ M. c nd = not determined. d 3,4-*O*isoprypylidene protected coupling product.

49 is favored in polar solvents and in reactions involving basic amines. This interesting elimination product **49** can be produced in high yield (95%) by reacting the bislactone **36** with 1 M NaOH in dioxane (Scheme 4).

A notable feature of the bislactone **36** is that the reaction rate for opening of the first lactone ring is considerably slower than the rate for opening of the second lactone ring. Thus only a minor amount of the monolactone product was traced during the reaction or was isolated from the product mixture when an excess of amine is used.

HIV-1 Protease Inhibition. (Table 1) HIV-1 protease was cloned and heterologously expressed in *Escherichia coli* as described elsewhere.26 The inhibitory effect of the synthesized compounds was initially determined with purified HIV-1 protease in a standardized spectrophotometric assay.27 The results are presented as IC_{50} values, i.e., the concentration of inhibitor result-

Scheme 4. A High Yielding Ring Opening Elimination Product

ing in 50% inhibition in this assay. For subsequent compounds and those that exhibited significant inhibitory effects, *K*ⁱ values were determined by a more sensitive fluorometric assay.²⁸

Structure-**Activity Relationship and X-ray Crystallographic Data.** For compounds **²³**-**²⁵** and **²⁷** each having lipophilic P2/P2′ amino acid methyl ester side chains, only compound **23** showed any enzyme inhibitory activity. A dramatic increase in inhibitory activity (IC_{50}) going from 5000 nM to 15 nM was obtained when the methyl ester of compound **23** was changed to the corresponding *N*-methyl amide (**28**). Thus duplication of one additional hydrogen bond between the C_2 -symmetric inhibitor and the HIV protease backbone results in this substantial increase in inhibitory activity.

To explore hydrophilic interactions to the enzyme backbone or to groups lining the hydrophobic S2/S2′ pockets, a number of amines each containing hydrogen bond donating or accepting functionalities were selected and coupled to give compounds **²⁶**, **²⁹**-**34**, and **⁴⁵**-**47**. However, these compounds exhibited only weak or no HIV-1 protease inhibitory activity. Efforts were then turned to optimizing the P2/P2′ side chains of the methyl amide **²⁸**. Compounds **³⁷**-**⁴²** and **⁴⁸** were prepared where enzyme inhibition data indicated that only lipophilic substituents at this position produced active inhibitors, K_i (nM) = 660, 140, 81, 2.3, and 0.9 for inhibitors **40** (Met), **37** (Phe), **42** (m-F-phenylGly), **41** (phenylGly), and **48** (Ile) where compounds **28** and **48** were the most potent in this series.

The X-ray crystal structure of HIV-1 protease complexed with the isoleucine inhibitor **48** (Figure 5) was determined.29 It reveals that one of the hydroxyls of the central diol points toward the active site Asp 25/

Figure 5. Schematic drawing showing the hydrogen bonds between the HIV-1 protease and the $\overline{C_2}$ -symmetric inhibitor **48**.

125 residues and is hydrogen bonded to both carboxyl oxygens. The other hydroxyl group points away from the active site but is still hydrogen bonded to one of the Asp residues. This difference in usage of the hydroxyls results in an asymmetric positioning of the inhibitor in the center of the protease. The P1/P1′ benzyloxy groups are positioned in the S1/S1′ pockets. The 1,6-dicarbonyl oxygens of the L-mannaric acid backbone and the amide nitrogens of the residues Ile 50/150 coordinate a structural water molecule in a distorted tetrahedral arrangement. The carbonyl oxygen of Ile (Figure 5) binds to the backbone amide nitrogens of Asp 29/129 in the S3/ S3′ site, and the nitrogen of the *N*-methyl amide binds to the Gly 48/148 carbonyl. The aliphatic Ile side chains are placed in the S2/S2′ pockets in close packing distances to Val 32/132. From this structural information it is apparent that the S2/S2′ pockets of the Ile inhibitor are not optimally filled up and that conformationally restricted side chains might induce improved fit to the S2/S2′ pockets.

1(*S*)-Amino-2(*R*)-hydroxyindan has successfully been incorporated in HIV-1 protease inhibitors by, for example, researchers at Merck (Figure 1, indinavir). As deduced from molecular modeling experiments, 1(*S*) amino-2(*R*)-hydroxyindan excellently fit with respect to filling the S2/S2′ subsite pockets and with respect to hydrogen bonding between the hydroxyl group and the protease backbone. Compound **43,** having the 1(*S*) amino-2(*R*)-hydroxyindan moiety, was thus synthesized and found to be twice as active as compound **28** (*K*ⁱ 0.2 nM). As expected the corresponding enantiomer of 1(*S*) amino-2(*R*)-hydroxyindan (**44**) exhibited no enzyme inhibition activity.

The X-ray crystal structure of HIV-1 protease complexed with inhibitor **43** (Figure 6) was determined, and reveled that both inhibitors **43** and **48** bind similarly.29 The hydroxyl group of 1(*S*)-amino-2(*R*)-hydroxyindan in inhibitor **43** binds to the backbone amide nitrogens of Asp 29/129 in the S3/S3′ site. The aromatic portion of the 1(*S*)-amino-2(*R*)-hydroxyindan groups in inhibitor **43** are positioned such that they truly form the intended expansion of the Ile groups, nicely filling out the S2/S2′ pockets.

Figure 6. Schematic drawing showing the hydrogen bonds between the HIV-1 protease and the $\overline{C_2}$ -symmetric inhibitor **43**.

Table 2. In Vitro Anti-HIV-1 Activity³⁰

O $\boldsymbol{\mathsf{A}}$ Ρh	Рh он i O Α OH O	
$A -$	Cmpd. no.	ED ₅₀ $(\mu g/mL)$
Ritonavir	$\mathbf{2}$	0.05
Indinavir	3	0.04
H N N H ő	28	0.9
H N	48	1.0
OН N	43	0.06

In Vitro Anti-HIV Activity. (Table 2) The anti-HIV activity was assayed by an HIV cytopathic assay in MT-4 cells where the effect was quantified using vital dye XTT. 30 The 50% inhibitory concentrations (ED₅₀) were calculated from the percent cytoprotection for individual compounds.

As can be seen in Table 2, compound **43** has an anti-HIV-1 activity in cell culture which is comparable to that of ritonavir and indenavir whereas compounds **28** and **48** are more than 10 times less active in the assay.

Conclusion

Application of the concept of C-terminal duplication with a carbohydrate, L-mannaric acid, as a new template for a peptidomimetic enabled the design of a number of potent HIV-1 protease inhibitors. One of the compounds, **43,** also demonstrated promising anti-HIV activity in cell culture. Two synthetic routes leading up to these inhibitors have been developed, one of which gives access to these inhibitors in just three chemical steps from commercially available starting materials.

The structure-activity relationships for these C_2 symmetric inhibitors have been rationalized with the aid of X-ray crystal structures. An IC_{50} going from 5000 nM to 15 nM was observed upon changing -OMe (**23**) into -NHMe (**28**), resulting in the net addition of one more hydrogen bond interaction from each NHMe group to the enzyme backbone. By sacrificing this important hydrogen bonding interaction but by optimizing binding to the S2/S2′ subsite pockets, an even more potent inhibitor was discovered (**43**). Further studies of this class of compounds are currently under way.

Experimental Section

Thin layer chromatography was performed using silica gel 60 F-254 (Merck) plates with detection by UV and/or charring with 8% sulfuric acid. Column chromatography was performed on silica gel (Matrix Silica Si 60A, 35-⁷⁰ *^µ*m, Amicon) eluting with CH3Cl-MeOH (gradient 5-20% MeOH) if not otherwise stated. Organic phases were dried over anhydrous sodium sulfate or magnesium sulfate. Concentrations were performed under reduced pressure. Optical rotations were recorded on a Perkin-Elmer 241 polarimeter. Infrared spectra were recorded on a Perkin-Elmer 1600 series FTIR instrument. NMR spectra were recorded on a JEOL GSX-270 instrument; 13C NMR 67 MHz and 1H NMR 270 MHz. Chemical shifts are reported in ppm (*δ*) downfield from tetramethylsilane in CDCl3, unless otherwise stated.

Accurate mass measurements were recorded on a JEOL SX 102 mass spectrometer/MS-MP7000 data system.

3,4-*O***-Isopropylidene-L-mannitol (12).** To a stirred and cooled (0 °C) solution of L-mannonic *γ*-lactone (20.0 g, 112 mmol) in dry MeOH (700 mL) was added lithium borohydride (4.80 g, 225 mmol) in four portions. The reaction mixture was stirred at room temperature for 30 min, cooled to 0 °C, and acidified with HOAc. After the mixture was stirred for 15 min, it was concentrated until dryness and the residue dissolved in MeOH (200 mL). Another portion of HOAc and MeOH was added and the acidic solution was again concentrated. Drying in a vacuum gave L-mannitol as a white foam which was suspended in dry acetone (600 mL) to which 2,2-dimethoxypropane (100 mL, 821 mmol) and camphorsulfonic acid (60.0 g, 260 mmol) were added. The reaction mixture was stirred overnight and concentrated. EtOAc (300 mL) was added, and the organic layer was washed with saturated aqueous $NaCO₃$ (300 mL), dried, and concentrated. Purification by column chromatography (toluene:EtOAc, 10:1) gave the triacetonide as a white solid (26.5 g, 87.6 mmol, 78%).

The triacetonide (25.1 g, 83.0 mmol) was dissolved in 70% HOAc (500 mL) and stirred at 40 °C for 1.5 h. The solution was concentrated, the residue extracted with acetone (Lmannitol remains as a solid residue), and the extract concentrated. The remaining traces of HOAc were removed by coevaporation with added toluene. The solid residue was recrystallized from warm acetone to give compound **12** (11.7 g, 53 mmol, 64%) as white crystals. 1H NMR was in agreement with that previously reported $9b$ (patent WO $93/07128$).

1,6-Di-*O***-***tert***-butyldimethylsilyl-3,4-***O***-isopropylidene-L-mannitol (13).** *tert*-Butyldimethylsilyl chloride (814 mg, 5.4 mmol) was added to a stirred solution of compound **12** (566 mg, 2.57 mmol) in pyridine (10 mL) containing (dimethylamino)pyridine (7 mg, 0.05 mmol). The mixture was stirred at 40 °C for 1.5 h and then concentrated to dryness by coevaporation with added toluene. The residue was dissolved in CH_2Cl_2 (30 mL) and washed with saturated aqueous NaCO₃, dried, concentrated, and purified by column chromatography (toluene:EtOAc, 10:1) to give **13** as a clear oil (1.10 g, 2.47 mmol, 96%). ¹³C NMR (CDCl₃): δ 5.2, 18.5, 26.0, 27.1, 64.6, 79.6, 109.4. Anal. $(C_{21}H_{46}O_6Si_2)$ C, H.

2,5-Di-*O***-benzyl-1,6-di-***O***-***tert***-butyldimethylsilyl-3,4-***O***isopropylidene-L-mannitol (14).** To a cold $(0 \degree C)$, stirred suspension of NaH (290 mg, 12.1 mmol) in dry THF (30 mL) under an atmosphere of argon was added **13** (2.27 g, 5.06 mmol) over 10 min. The ice bath was removed, and the resulting tan solution was stirred at room temperature for 15 min. Tetrabutylammonium iodide (15 mg) and benzyl bromide (1.32 mL, 11.1 mmol) were added, and the reaction mixture was stirred at room temperature for 15 h. $H₂O$ (3 mL) was added, and the resulting mixture was concentrated. The residue was taken up in Et₂O (40 mL) and washed with H_2O $(3 \times 40 \text{ mL})$. The combined organic extracts were dried, concentrated, and purified by column chromatography (light petroleum ether-EtOAc, 17:1) to give compound **¹⁴** as a clear oil (2.43 g, 3.85 mmol, 76%). 13C NMR (CDCl3): *δ* 5.4, 18.2, 25.9, 27.2, 63.8, 73.1, 78.3, 81.1, 109.5, 127.7, 128.1, 138.8. Anal. $(C_{35}H_{58}O_6Si_2)$ H,C.

2,5-Di-*O***-benzyl-3,4-***O***-isopropylidene-L-mannitol (15).** Compound **14** (2.10 g, 3.3 mmol) was dissolved in THF (15 mL), a solution of tetrabutylammonium fluoride in THF (7.2 mL, 1.1 M) was added, and the reaction mixture was stirred at room temperature for 1 h. Most of the solvent was removed under reduced pressure, and the residue was flashed through a short column of silica gel with $CH₃CN$ as eluent, concentrated, and purified by column chromatography (light petroleum ether-EtOAc, 1:1) to give compound **¹⁵** as a clear oil $(1.29 \text{ mg}, 3.2 \text{ mmol}, 98\%)$ which solidified upon standing. 13 C NMR (CDCl₃): δ 27.3, 61.4, 80.0, 110.1, 128.1, 128.7, 137.9. Anal. $(C_{23}H_{30}O_6)$ C, H.

2,5-Di-*O***-benzyl-3,4-***O***-isopropylidene-L-mannaric acid (16).** To a stirred solution of compound **15** (1.080 g, 2.68 mmol) in CH_2Cl_2 (36 mL) was added TEMPO (2,2,6,6-tetramethyl-1-piperidinyloxy, free radical) (22 mg, 0.14 mmol) and a solution of saturated aqueous $NaCO₃$ (11 mL), KBr (59 mg, 0.50 mmol), and tetrabutylammonium bromide (90 mg, 0.28 mmol). The mixture was cooled to 0 °C, and a solution of sodium hypochlorite (17.4 mL, 1.2 M), saturated aqueous $NaCO₃$ (5.9 mL), and brine (11.8 mL) was added over 45 min. After the mixture was stirred at 0 °C for an additional 45 min, the organic layer was separated and washed with H₂O (3 \times 20 mL). The combined aqueous phases were acidified with 4 M HCl, extracted with EtOAc $(3 \times 20$ mL), dried, and concentrated to give compound **16** as a white solid (775 mg, 1.80 mmol, 67%). 13C NMR (DMSO-*d*6): *δ* 27.0, 71.7, 78.4, 79.1, 110.0, 127.6, 128.3, 137.5, 170.6. A small portion was recrystallized from EtOAc and light petroleum ether to give the dicarboxylic acid as white crystals which were subjected to elementary analysis. Anal. $(C_{23}H_{26}O_8)$ C, H.

Procedure for Preparation of Compounds 17-**21. Method I.** Compound **¹⁶** (200-300 mg, 0.46-0.70 mmol), amino acid derivative $(2.2 \text{ equity}, 1.01-1.54 \text{ mmol})$, and 1 -hydroxybenzotriazole hydrate (3.0 equiv, 186-284 mg, 1.38-2.10 mol) were dissolved in $6-8$ mL CH₂Cl₂-THF (2:1) under an atmosphere of argon. The pH of the stirred reaction mixture was adjusted to 7.5 by dropwise addition of triethylamine. The temperature was lowered to 0 °C, and 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (2.4 equiv, 211-³²² mg, 0.96-1.68 mmol) was added in one portion. After 1 h, the temperature was raised to room temperature, and the reaction mixture was stirred for $2-4$ h, depending on the amino acid derivative used. CH_2Cl_2 (20 mL) was added, and the organic layer was washed with saturated aqueous NaCO₃. The organic layer was separated, dried, and concentrated. Column chromatography (toluene-EtOAc, 3:1-4:1) furnished compounds **¹⁷**-**21**. Details about yields and amino acid derivatives used are listed in Table 1.

*N***1,***N***6-Di[(1***S***)-2-methyl-1-(methoxycarbonyl)propyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxy-3,4-***O***-isopropylidenehexanediamide (17).** 13C NMR (CDCl3): *δ*

17.8, 19.0, 27.0, 31.2, 52.0, 56.7, 73.9, 77.6, 79.4, 110.4, 128.2, 128.3, 136.8, 169.0, 171.8. Anal. (C₃₅H₄₈N₂O₁₀·1H₂O) C, H, N.

*N***1,***N***6-Di[(1***S***)-2-phenyl-1-(methoxycarbonyl)ethyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxy-3,4-***O***-isopropylidenehexanediamide (18).** 13C NMR (CDCl3): *δ* 27.0, 37.9, 52.1, 52.6, 73.6, 78.1, 79.8, 110.6, 128.4, 128.5, 128.7, 129.2, 135.6, 136.5, 169.0, 171.6. Anal. $(C_{43}H_{48}N_2O_{10} \cdot 0.5H_2O)$ C, H, N.

*N***1,***N***6-Di[(1***S***)-2-methyl-1-(benzyloxycarbonyl)propyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxy-3,4-***O***-isopropylidenehexanediamide (19).** 13C NMR (CDCl3): *δ* 17.67, 19.0, 27.0, 31.3, 56.7, 66.9, 73.9, 77.5, 79.5, 110,4, 128.2, 128.3, 128.6, 129.5, 135.5, 136.9, 169.1, 171.2. Anal. $(C_{47}H_{56}N_3 O_{10}$ \cdot 0.5H₂O) C, H, N.

*N***1,***N***6-Di[(1***S***)-2-methyl-1-(hydroxymethyl)propyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxy-3,4-***O***-isopropylidenehexanediamide (20).** Compound **20** was directly subjected to deprotection. 13C NMR (CDCl3): *δ* 18.2, 19.4, 27.3, 30.2, 56.9, 63.3, 75.1, 77.5, 79.3, 110.4, 128.3, 128.8, 129.0, 136.7, 169.6.

*N***1,***N***6-Di[(1***S***)-3-methyl-1-(methoxycarbonyl)butyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxy-3,4-***O***-isopropylidenehexanediamide (21).** Compound **21** was directly subjected to deprotection. 13C NMR (CDCl3): *δ* 21.9, 23.0, 24.9, 27.1, 41.7, 50.3, 52.3, 74.0, 77.8, 79.6, 110,5; 128.4, 128.7, 129.1, 136.9, 169.2, 173.0.

*N***1,***N***6-Disuccinimidyl-(2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)- 3,4-dihydroxyhexanediamide (22).** A mixture of compound **16** (1.01 g, 2.35 mmol), *N*,*N*-disuccinimidyl carbonate (2.41 g, 9.4 mmol) and pyridine (1.15 mL, 14.1 mmol) in CH3CN (15 mL) was stirred at ambient temperature overnight. The mixture was concentrated and the residue dissolved in EtOAc. The organic phase was washed with H2O, dried, and concentrated, and the residue was purified by column chromathograpy to give **22** (1.29 g, 2.07 mmol, 88%) as a white foam. A small portion was crystallized from 2-propanol. $[\alpha]D = +53.8^{\circ}$ (*c* = 0.935, CHCl3). 1H NMR (CDCl3): *δ* 1.4 (s, 6H), 2.8 (s, 8H), 4.4-4.65 (m, 6H), 4.85 (d, 2H), 7.2-7.4 (m, 10H). 13C NMR (CDCl3): *δ* 25.6, 27.0, 73.0, 77.0, 77.8, 112.2, 128.2, 128.5, 136.1, 165.5, 168.4. Anal. (C31H32N2O12) C, H, N.

Procedure for Preparation of Compounds 23-27.
Method I. Compounds 17-21 were dissolved in MeOH (15) **Method I.** Compounds **¹⁷**-**²¹** were dissolved in MeOH (15 mL) acidified with HCl (4%, w:w) and stirred at room temperature for 2 h. The clear solution was concentrated and purified by column chromatography (toluene-EtOAc, 1:1; CH₂- Cl_2 –MeOH, 95:5 for compound **26**) to give compounds **23–27** in 70-74% yield (see Table 1).

*N***1,***N***6-Di[(1***S***)-2-methyl-1-(methoxycarbonyl)propyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (23).** ¹³C NMR (CDCl₃): δ 17.6, 19.0, 31.3, 52.1, 56.6, 71.6, 74.2, 78.0, 128.3, 128.5, 136.4, 171.7, 172.1. $[\alpha]_{D} =$ $+20.53^{\circ}$ ($c = 1.13$, CHCl₃). Anal. (C₃₂H₄₄N₂O₁₀·0.5H₂O) C, H, N.

*N***1,***N***6-Di[(1***S***)-2-phenyl-1-(methoxycarbonyl)ethyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (24).** $[\alpha]_D = +44.21$ ° ($c = 1.07$, CHCl₃). ¹³C NMR (CDCl3): *δ* 37.7, 52.6, 53.0, 70.9, 74.0, 79.1, 128.3, 128.7, 128.8, 129.2, 135.8, 136.8, 171.5, 172.3. Anal. $(C_{40}H_{44}N_2O_{10} \cdot 1H_2O)$ C, H, N.

*N***1,***N***6-Di[(1***S***)-2-methyl-1-(benzyloxycarbonyl)propyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (25).** $[\alpha]_D = +16.11^{\circ}$ ($c = 0.95$, CHCl₃). ¹³C NMR (CDCl3): *δ* 17.5, 18.9, 31.0, 56.9, 67.0, 70.8, 74.0, 78.4, 128.4, 128.6, 128.7, 129.6, 135.3, 136.8, 171.0, 172.6. Anal. (C₄₄H₅₂N₂-O10'1H2O) C, H, N.

*N***1,***N***6-Di[(1***S***)-2-methyl-1-(hydroxymethyl)propyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (26).** ¹³C NMR (CDCl₃): δ 18.9, 19.6, 29.0, 56.9, 62.9, 71.8, 73.2, 81.1, 128.1, 128.3, 128.6, 136.8, 171.7. Anal. $(C_{30}H_{44}N_2O_8 \cdot 1H_2O)$ C, H, N.

*N***1,***N***6-Di[(1***S***)-3-methyl-1-(methoxycarbonyl)butyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (27).** ¹³C NMR (CDCl₃): δ 21.7, 23.0, 24.9, 27.1, 41.7, 50.4, 52.3, 70.9, 74.0, 78.8, 128.2, 128.3, 128.7, 136.9, 172.4, 172.7. Anal. (C₃₄H₄₈N₂O₁₀) C, H, N.

Valine Methylamide. To a cooled $(-15 \degree C)$ solution of Z-Val-OSu (348 mg, 1 mmol) in dry THF (5 mL) was added methylamine (2 mmol, 2 M solution in THF). The reaction mixture was brought to room temperature and stirred for 4 h. The *N*-hydroxysuccinimide was filtered off and the residue concentrated, dissolved in EtOAc (25 mL), washed with NaHCO₃ (2×15 mL), dried, and concentrated to give the crude methylamide which was crystallized from EtOAc. The product was dissolved in EtOAc and hydrogenated over Pd/C for 6 h. The catalyst was removed and the residue concentrated to give the desired methylamide in 78% yield (102 mg, 0.78 mmol). Spectral data were in agreement with those previously reported.19

*N***1,***N***6-Di[(1***S***)-2-methyl-1-(methylcarbamoyl)propyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (28).** Compound **16** (181 mg, 0.42 mmol), valine methylamide (121 mg, 0.93 mmol), and 1-hydroxybenzotriazole hydrate (170 mg, 1.26 mmol) were dissolved in 6 mL of CH_{2} - Cl_2 -THF (2:1) under an atmosphere of argon. The pH of the stirred reaction mixture was adjusted to 7.2 by dropwise addition of triethylamine. The temperature was lowered to 0 °C, and 1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride (191 mg, 1.0 mmol) was added in one portion. After 1 h, the temperature was raised to room temperature and the reaction mixture was stirred 4 h. CH_2Cl_2 (20 mL) was added, and the organic layer was washed with saturated aqueous NaCO₃. The organic layer was separated, dried, and concentrated. The residue was dissolved in MeOH (15 mL) acidified with HCl (4%, w:w) and stirred at room temperature for 2 h. The clear solution was concentrated and purified by column chromatography (toluene-EtOAc, 1:1) to give **²⁸** in 88% yield. $[\alpha]_D = -6.27^{\circ}$ ($c = 1.03$, CHCl₃). ¹³C NMR (CDCl₃-CD₃OD, 7:3): *δ* 17.5, 19.6, 26.0, 30.2, 58.5, 72.1, 73.2, 80.5, 128.4, 128.6, 137.2, 172.1, 172.4. Anal. $(C_{32}H_{46}N_4O_8.1H_2O)$ C, H, N.

General Method for the Preparation of Compounds ²⁹-**31. Method II.** The l-mannaric acid **¹⁶** (1.0 equiv), the amine (2.2 equiv), and HOBT (3.0 equiv) were dissolved in 2:1 CH_2Cl_2 -THF with stirring under argon. The pH was adjusted to 7.5 by dropwise addition of triethylamine, and the temperature was adjusted to 0 °C. EDC (2.4 equiv) was added, and stirring was continued for 1 h at 0 °C and then for 16 h at ambient temperature. The reaction mixture was diluted with CH_2Cl_2 (2 x volume), washed with saturated aqueous NaHCO₃, dried, and concentrated, and the residue was purified by column chromatography. The purified l-mannaric amides were dissolved in 5 mL of 4% HCl in MeOH, and the reaction mixture was stirred at ambient temperature until TLC indicated complete reaction. The mixture was partitioned between CH_2Cl_2 and saturated aqueous NaHCO₃, dried, concentrated, and purified by column chromatography to give the target compounds **²⁹**-**31**.

*N***1,***N***6-Di(2-pyridylmethyl)-(2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (29).** The title compound was prepared in 29% yield (50 mg, 0.088 mmol) according to method II, using 2-aminomethyl pyridine $[\alpha]_D =$ $+21.7^{\circ}$ ($c = 1.34$, CHCl₃). ¹³C NMR (CDCl₃): δ 43.9, 71.3, 73.7, 79.5, 121.9, 122.4, 128.1, 128.3, 128.5, 137.0, 148.9, 156.1, 172.4. Anal. $(C_{32}H_{34}N_4O_6 \cdot 0.5H_2O)$ C, H, N.

*N***1,***N***6-Di(3-pyridylmethyl)-(2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4- dihydroxyhexanediamide (30).** The title compound was prepared in 30% yield **30** (62 mg, 0.109 mmol) according to method II, using 3-aminomethyl pyridine. $[\alpha]_D$
= +49.2° $(c=1.34$ CHCl³ \rightarrow ¹³C NMR (CD₂OD); δ 40.5, 40.6 $= +49.2^{\circ}$ (*c* = 1.34, CHCl₃). ¹³C NMR (CD₃OD): *δ* 40.5, 40.6, 71 2 73 3 80 3 124 2 128 4 128 7 136 4 137 1 148 0 71.2, 73.3, 80.3, 124.2, 128.4, 128.7, 134.7, 136.4, 137.1, 148.0, 148.5, 172.6, 172.7. Anal. (C₃₂H₃₄N₄O₆) C, H, N.

*N***1,***N***6-Di(4-pyridylmethyl)-(2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (31).** The title compound was prepared in 21% (94 mg, 0.165 mmol) according to method II, using 4-aminomethyl pyridine. $[\alpha]_D = +31.4^{\circ}$ (*c* = 0.36, MeOH). 13C NMR (CDCl3): *δ* 41.8, 71.5, 73.8, 79.8, 122.2, 128.3, 128.5, 128.7, 136.6, 147.2, 149.6, 172.3. Anal. (C₃₂H₃₄N₄- O_6 \cdot 0.5H₂O) C, H, N.

General Method for the Preparation of Compounds ³²-**34. Method III.** The succinimidyl diester **²²** (1.0 equiv) and the amine (2.5 equiv) were dissolved in dichloroethane. The mixture was stirred for 16 h under argon at 65 °C. After concentration the mixture was dissolved in CH_2Cl_2 and washed with saturated aqueous NaHCO₃, dried, and concentrated, and the residue was subjected to column chromathograpy. The purified l-mannaric amides were dissolved in 5 mL of 4% HCl in MeOH, and the reaction mixture was stirred at ambient temperature until TLC indicated complete reaction. The mixture was partitioned between CH_2Cl_2 and saturated aqueous NaHCO₃ ($2\times$), dried, concentrated, and purified by column chromatography to give title compounds **³²**-**34**.

*N***1,***N***6-Di(3-hydroxy-2-methylphenyl)-(2***R***,3***R***,4***R***,5***R***)- 2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (32).** The title compound was prepared in 25% yield (40 mg, 0.067 mmol) according to method III, using 3-amino-*o*-cresol. $[\alpha]_D = +37.8^\circ$ $(c = 0.54, \text{MeOH})$. ¹³C NMR (CD₃OD and CDCl₃): δ 10.0, 71.5, 73.9, 80.4, 112.8, 115.4, 118.0, 126.6, 128.6, 128.9, 135.8, 137.1, 155.7, 170.8. HRMS calcd for $C_{34}H_{36}N_2O_8$ 600.2472, found 600.2508.

*N***1,***N***6-Di(2-hydroxy-4-methylphenyl)-(2***R***,3***R***,4***R***,5***R***)- 2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (33).** The title compound was prepared in 32% yield (75 mg, 0.13 mmol) according to method III, using 6-amino-*m*-cresol. $[\alpha]_D = +46.2^{\circ}$ (*^c*) 0.75, MeOH). 13C NMR (CD3OD): *^δ* 21.2, 72.4, 74.1, 81.7, 116.9, 121.0, 122.4, 124.2, 129.0, 129.4, 129.5, 136.4, 138.6, 148.9, 171.7. Anal. (C₃₄H₃₆N₂O₈) C, H, N.

*N***1,***N***6-Di(4-hydroxy-2-methylphenyl)-(2***R***,3***R***,4***R***,5***R***)- 2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (34).** The title compound was prepared in 38% yield (96 mg, 0.16 mmol) according to method III, using 4-amino-3-methylfenol. $[\alpha]_D =$ $+30.3^{\circ}$ ($c = 0.75$, DMSO). ¹³C NMR (DMSO- d_6): δ 18.0, 70.0, 71.5, 80.2, 112.6, 116.6, 127.2, 127.5, 127.6, 127.8, 128.2, 134.1, 138.0, 155.1, 169.9. Anal. (C34H36N2O8) C, H, N.

L-Mannaro-1,4:3:6-di-*γ***-lactone (35).** L-Mannonic *γ*-lactone (11) (1.0 equiv, 3.0 g, 17.0 mmol), HNO₃ (9 mL, concentrated), and H_2O (1.7 mL) were heated to 85 °C under stirring with the evolution of nitrous gases. The reaction mixture was refluxed for 16 h and concentrated twice with addition of H_2O (30 mL). The residue was lyophilized from $H₂O$ (40 mL). The resulting white solid was suspended in ethanol (5 mL) and diethyl ether (30 mL). After filtration and drying **35** was obtained as a white solid (1.76 g, 10.1 mmol, 60%). $[\alpha]_D =$ -158.81° ($c = 1.18$, MeOH). ¹H NMR (DMSO- d_6): δ 4.75 (d, 2H), 5.1 (d, 2H), 6.4 (broad s, 2H). 13C NMR (DMSO-*d*6): *δ* 69.2, 75.9, 174.3. IR (KBr): 1799.4 cm⁻¹. Anal. (C₆H₆O₆) C, H.

2,5-Di-*O***-benzyl-L-mannaro-1,4:3,6-di-***γ***-lactone (36).** To a stirred solution of L-mannaro-1,4:6,3-di-*γ*-lactone (300 mg, 1.72 mmol) and benzyl-2,2,2-trichloroacetimidate (0.96 mL, 5.17 mmol) in dry dioxane was added trifloromethanesulfonic acid (240 μ L) dropwise under nitrogen. Within 1-2 h the color changed to red-brown. After an additional 30 min the reaction mixture was filtered through 2 cm of $NaCO₃$ on 2 cm of silica in a glass filter-funnel and subsequently evaporated under reduced pressure. Warm diethyl ether was added to the crude product, the mixture was stirred for 1 min, and the diethyl ether was thereafter decanted. Recrystalization from CH_2Cl_2 gave white crystals of 2,5-di-*O*-benzyl-L-mannaro-1,4:3,6-di-
 γ -lactone (439.5 mg, 72%). [α]_D = -125.58° (*c* = 1.04, CH₂*γ*-lactone (439.5 mg, 72%). [α]_D = -125.58° (*c* = 1.04, CH₂-
Cl₉) IR (KBr) 1785 cm⁻¹ Mp 184–186 °C ⁻¹³C NMR (DMSO-Cl₂). IR (KBr) 1785 cm⁻¹. Mp 184–186 °C. ¹³C NMR (DMSO-
d₀): δ 72 04 74 39 74 82 127 97 128 04 128 4 136 92 171 76 *d*6): *δ* 72.04, 74.39, 74.82, 127.97, 128.04, 128.4, 136.92, 171.76. Anal. $(C_{20}H_{18}O_6)$ C, H.

General Method for the Preparation of Compounds 28, 37-**40. Method IV.** Bislactone **³⁶** (40 mg, 0.11 mmol) was dissolved in CH_2Cl_2 (4 mL). Six equivalents of the amino acid derivative was added to the solution, and the mixture was stirred at reflux for 16 h. The solvent was removed under reduced pressure and the crude product purified by silica gel chromatography.

*N***1,***N***6-Di[(1***S***)-2-methyl-1-(methylcarbamoyl)propyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (28).** The title compound was prepared using valine methylamide 19 according to method IV using $\rm CH_2Cl_2\rm-MeOH$ (20: 1) as eluent to give **28** (48.6 mg, 70%).

*N***1,***N***6-Di[(1***S***)-2-phenyl-1-(methylcarbamoyl)ethyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (37).** The title compound was prepared using phenylalanine methylamide³¹ according to method IV using CH_2Cl_2- MeOH (20:1) as eluent to give product **37** (60.9 mg, 76%). $\left[\alpha\right]_D$ = +5.59° (c = 1.02, CHCl₃); IR (KBr): 3324, 1650, 1525 cm⁻¹. ¹³C NMR (CDCl₃): δ 26.20, 37.02, 54.03, 72.83, 73.67, 81.87, 127.07, 127.81, 128.42, 128.65, 128.78, 129.00, 136.38, 134.98, 170.00, 172.06. Anal. $(C_{40}H_{46}N_4O_8)$ C, H, N.

*N***1,***N***6-Di[(1***S***)-2-(***p***-hydroxy)phenyl-1-(methylcarbamoyl)ethyl]-(2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (38).** The title compound was prepared using tyrosine methylamide³¹ according to method IV using CH_2Cl_2-MeOH (9:1) as eluent to give product **38** (60.0 mg, 60%). $[\alpha]_D = +7.13^\circ$ ($c = 0.87$, EtOH). IR (KBr): 3320, 1651, 1514 cm-1. 13C NMR (CD3OD): *δ* 26.13, 37.07, 55.32, 72.86, 72.93, 81.13, 116.14, 128.83, 129.15, 130.93, 138.32, 157.18, 172.92, 173.59. Anal. $(C_{40}H_{46}N_4O_{10})$ C, H, N.

*N***1,***N***6-Di[(1***S***,2***R***)-2-hydroxy-1-(methylcarbamoyl)propyl]-(2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (39).** The title compound was prepared according to method IV using threonine methylamide using CH_2Cl_2- MeOH (9:1) to give product **39** (42.9 mg, 61%). [α]_D = -3.24°
(c = 1.02 MeOH) IR (KBr): 3393 1651 1518 cm^{-1 13}C NMR (*c* = 1.02, MeOH). IR (KBr): 3393, 1651, 1518 cm⁻¹. ¹³C NMR
(CD₂OD): δ 20 59 26 18 59 53 67 09 73 13 81 02 128 90 (CD3OD): *δ* 20.59, 26.18, 59.53, 67.09, 73.13, 81.02, 128.90, 129.08, 129.24, 138.39, 172.69, 173.21. Anal. (C₃₀H₄₂N₄O₁₀) C, H, N.

*N***1,***N***6-Di[(1***S***)-3-thiomethyl-1-(methylcarbamoyl)propyl]-(2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (40).** The title compound was prepared using methionine methylamide³² according to method IV using $CH₂$ - Cl_2-MeOH (20:1) as eluent to give product **40 (**55.0 mg, 72%). $[\alpha]_D = -13.84^{\circ}$ ($c = 1.51$, MeOH). IR (KBr): 3303, 1636, 1541 cm-1. 13C NMR (CD3OD): *δ* 15.37, 26.58, 31.42, 32.21, 53.73, 72.72, 73.64, 81.55, 129.16, 129.41, 129.60, 138.86, 174.02, 174.22. Anal. $(C_{32}H_{46}N_4S_2O_8)$ C, H, N.

General Method for the Preparation of Compounds ⁴¹-**48. Method IV.** A mixture of the amine (2.2-6 equiv) in specified solvent was heated at 40-70 °C with stirring. The benzylated bislactone **36** (1.0 equiv) was added, and the mixture was stirred for $1-16$ h at elevated temperature. The solvent was evaporated, and the crude product was purified by silica gel column chromatography and/or crystallization to give the target compounds **⁴¹**-**48**.

L-Phenylglycine Methyl Amide. Z-PhGly-OSu (1 equiv, 2.01 g, 5.26 mmol) was suspended in THF (20 mL) under argon at -10 °C. Methylamine (1.6 equiv, 4.2 mL as a 2.0 M solution in THF, 8.4 mmol) was added dropwise during 20 min. Stirring was continued at -10 °C for 2 h and at room temperature for 1.5 h. The mixture was filtered through Celite, concentrated, and partitioned between $CHCl₃$, $H₂O$, and NH4Cl, dried, and concentrated. Crystallization from EtOAc gave Z-L-phenylglycine methyl amide **(**649 mg, 2.18 mmol, $\overline{42\%}$) as a white solid. ¹³C NMR (CD₃OD and CDCl₃): δ 26.3, 26.5, 58.7, 67.2, 127.2, 128.0, 128.3, 128.5, 128.6, 129.0, 136.2, 138.1, 156.3, 171.3.

*Z***-L-Phenylglycine Methyl Amide** (1.0 equiv, 607 mg, 2.03 mmol) was dissolved in EtOAc (12 mL). Pd/C (44 mg) was added, and the suspension was stirred under hydrogen for 21 h, filtered through Celite, and concentrated to give L-phenylglycine methyl amide (274 mg, 1.67 mmol, 77%) as a yellow oil which was used in the next step without further purification. ¹³C NMR (CD₃OD): δ 26.4, 59.8, 128.2, 129.3, 129.8, 140.9, 174.9.

*N***1,***N***6-Di[(1***S***)-1-phenyl-1-(methylcarbamoyl)methyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (41).** The title compound was prepared in 31% yield (89 mg, 0.130 mmol) according to method IV, using l-phenylglycine methyl amide (274 mg, 1.67 mmol) in $\rm CH_3CN$ (1 mL) at 65 °C for 14 h. $[\alpha]_D = +75.7^\circ$ ($c = 0.39$, CH₃OH). ¹³C NMR
(CD2OD and CDCl2): δ 26.3, 26.4, 57.1, 71.8, 73.4, 80.3, 127.4 (CD3OD and CDCl3): *δ* 26.3, 26.4, 57.1, 71.8, 73.4, 80.3, 127.4, 128.3, 128.4, 128.5, 128.6, 129.0, 136.5, 137.2, 170.3, 171.3. Anal. Calcd for C₃₈H₄₂N₄O₈: C, 66.85; H, 6.20; N, 8.21. Found: C, 66.74; H, 6.34; N, 8.12.

*N***1,***N***6-Di[1-(3-fluorophenyl)-1-(methylcarbamoyl)methyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (42).** The title compound was prepared in 20% yield (49 mg, 0.068 mmol) according to method IV, using 3-fluorophenylglycine methyl amide (220 mg, 1.21 mmol) in CH₃CN (1 mL) at 70 °C for 16 h. ¹³C NMR (CD₃OD and CDCl3): *δ* 26.5, 57.4, 57.6, 71.7, 71.9, 72.2, 73.6, 73.7, 80.2, 80.3, 80.9, 115.0, 115.2, 115.4, 115.5, 115.7, 116.0, 124.0, 124.3, 128.7, 128.9, 129.2, 131.2, 131.3, 138.1, 140.9, 162.1, 165.7, 171.7, 172.8, 173.0. Anal. (C₃₈H₄₀N₄O₈F₂) C, H, N.

*N***1,***N***6-Di[(2***R***)-hydroxy-1(***S***)-indanyl]-(2***R***,3***R***,4***R***,5***R***)- 2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (43).** The title compound was prepared according to method IV, using (1*S*,2*R*)-(-)-*cis*-1-amino-2-indanol (0.915 g, 6.2 mmol) in CHCl3 (12 mL) at 45 °C for 4. The reaction mixture was concentrated and partitioned between CHCl₃ and saturated aqueous NH₄-Cl and H_2O . The organic layer was separated, dried (MgSO₄), and concentrated. Crystallization from MeOH gave **43 (**0.65 g, 0.996 mmol, 35%). [α]_D = -20.7° (*c* = 0.68, CHCl₃). ¹³C
NMR (CDCl₃): *δ* 39.2, 57.8, 71.6, 72.5, 73.5, 81.5, 124.0, 125.3, 127.0, 128.2, 128.3, 128.6, 136.7, 139.8, 140.8, 171.6. Anal. $(C_{38}H_{40}N_2O_8)$ C, H, N.

*N***1,***N***6-Di[(2***R***)-hydroxy-1(***S***)-indanyl]-(2***R***,3***R***,4***R***,5***R***)- 2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (44).** The title compound was prepared in 22% yield (60 mg, 0.092 mmol) according to method IV using (1*R*,2*S*)-(+)-*cis*-1-amino-2-indanol (370 mg, 2.5 mmol) in CH3CN (1 mL) at 70 °C for 16 h. $[\alpha]_D = +3.2^{\circ}$ ($c = 0.53$, CH₃OH). ¹³C NMR (CD₃OD and CDCl3): *δ* 40.1, 57.9, 71.6, 73.1, 73.5, 80.4, 124.8, 125.6, 127.5, 128.6, 128.8, 128.9, 137.6, 140.9, 173.2. Anal. (C₃₈H₄₀N₂O₈) C, H, N.

*N***1,***N***6-Di(2-hydroxyethyl)-(2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (45).** The title compound was prepared in 49% yield (105 mg, 0.296 mmol) according to method IV using 2-hydroxyethylamine (43 *µ*L, 0.78 mmol) in CHCl₃ (2 mL) at 40 °C for 1 h. $[\alpha]_D = +64.0$ ° (*c* $= 0.69$, CHCl₃). ¹³C NMR (CD₃OD): δ 42.5, 61.4, 71.6, 73.6, 81.2, 128.8, 129.2, 138.5, 174.0. Anal. (C₂₄H₃₂N₂O₈) C, H, N.

5-Aminomethyl Thiazole. 5-Hydroxymethyl thiazole (1.0 equiv, 315 mg, 2.74 mmol), triphenyl phosphine (1.4 equiv, 995 mg, 3.79 mmol), and $LiN₃$ (6.3 equiv, 855 mg, 17.5 mmol) were suspended in DMF (15 mL). CBr_4 (1.4 equiv, 1.25 g, 3.78) mmol) was added, and the reaction mixture was stirred at room temperature for 23 h after which MeOH (3 mL) was added. The mixture was partitioned between toluene and H_2O $(4\times)$, dried, and concentrated. The residue was purified using column chromathograpy eluting with toluene-EtOAc (5:1) to give 5-azidomethyl thiazole (297 mg, 2.12 mmol, 77%) as a yellow viscous oil. 1H NMR (CDCl3): *δ* 4.58, 7.74, 8.84. 13C NMR (CDCl₃): δ 46.3, 132.4, 142.7, 154.3. Anal. (C₄H₄N₄S) C, H, N. 5-Azidomethyl thiazole (1.0 equiv, 229 mg, 1.63 mmol) was dissolved in ethanol (20 mL), and Pd/C (0.2 g) was added. The suspension was stirred under hydrogen atmosphere for 20 h, filtered through Celite, and concentrated to give 5-aminomethyl thiazole (138 mg, 1.21 mmol, 74%) as a yellow syrup which was used without further purification. 1H NMR (CD₃OD): δ 4.04, 4.71, 7.76, 8.89. ¹³C NMR (CD₃OD): *δ* 38.5, 140.9, 141.7, 154.6.

*N***1,***N***6-Di(5-thiazolylmethyl)-(2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanediamide (46).** The title compound was prepared in 65% yield (106 mg, 0.182 mmol) according to method IV using 5-aminomethyl thiazole (138 mg, 1.21 mmol) in CH₃CN (1 mL) at 70 °C for 1 h. $[\alpha]_D = +29.0^{\circ}$ $(c = 0.38, CH_3OH)$. ¹³C NMR (CD₃OD and CDCl₃): δ 35.5, 71.3, 73.4, 80.6, 128.6, 128.9, 129.0, 137.7, 137.8, 141.6, 155.0, 173.4. Anal. $(C_{28}H_{30}N_4O_6S_2)$ C, H, N.

*N***1,***N***6-Di(2-chloro-6-fluorobenzyl)-(2***R***,3***R***,4***R***,5***R***)-2,5 di(benzyloxy)-3,4-dihydroxyhexanediamide (47).** The title compound was prepared in 59% yield (114 mg, 0.169 mmol) according to method IV using 2-chloro-6-fluorobenzylamine (282 mg, 1.77 mmol) in CH3CN (1 mL) at 70 °C for 21 h. $[\alpha]_D = +15.6^{\circ}$ ($c = 0.93$, CHCl₃). ¹³C NMR (CD₃OD and CDCl3): *δ* 34.8, 71.0, 73.7, 80.0, 114.5, 114.9, 123.3, 123.6, 125.8, 125.9, 128.5, 128.6, 128.8, 130.3, 130.4, 135.9, 137.2, 160.2, 163.9, 172.3, 172.4. Anal. $(C_{34}H_{32}N_2O_6Cl_2F_2)$ C, H, N.

*N***1,***N***6-Di[(1***S***,2***S***)-2-methyl-1-(methylcarbamoyl)butyl]- (2***R***,3***R***,4***R***,5***R***)-2,5-di(benzyloxy)-3,4-dihydroxyhexanedi**amide (48). Prepared using isolucine methylamide³³ according to method IV using chloroform—methanol (20:1) as eluent
to give (34.4 mg, 46%) = $(0.6 - 6.8^\circ)$ (c = 0.50 CHCl) = IR to give (34.4 mg, 46%). [α]_D = -6.8° (*c* = 0.50, CHCl₃). IR
(KBr): 3324, 1650, 1541 cm⁻¹. ¹³C (CD₃OD): *δ* 11.67, 16.07, 25.13, 26.34, 37.20, 58.46, 72.26, 73.39, 80.84, 128.72, 129.13, 137.72, 172.68, 173.12. Anal. $(C_{34}H_{50}N_4O_8)$ C, H, N.

(5*R***)-5-[(***R***)-(Benzyloxy)-(hydroxycarbonyl)methyl]-2 benzyloxy-2(5***H***)-furanone (49).** Bislactone **36** (40 mg, 0.113 mmol) was dissolved in dioxane (6 mL). NaOH (1 M, 2 mL) was added, and the reaction mixture was stirred for 5 min. The solvent was removed under reduced pressure, and the crude product was dissolved in saturated $NaCO₃$ (20 mL). The solution was extracted with CH_2Cl_2 (3 \times 20 mL). The combined organic phase was dried with MgSO₄, filtered, and concentrated under reduced pressure to give product **49** (38 mg, 95%). [α]_D = -16.8° (*c* = 1.06, MeOH). IR (KBr): 3428.7,
1781.3, 1651.0, 1590.2 cm⁻¹. ¹H NMR (CD₃OD): δ4.43 (d, *J* $=$ 3.9, 1H), 4.49 (d, $J = 11.6$, 1H), 4.74 (d, $J = 11.6$, 1H), 4.95 $(s, 2H), 5.32$ (dd, $J = 2.3, J = 3.8, 1H), 6.31$ (d, $J = 2.3, 1H$), 7.32 (m, 10H). 13C NMR (CD3OD): *δ* 73.74, 74.19, 78.27, 79.60, 115.75, 128.72, 128.95, 129.19, 129.37, 136.43, 138.15, 147.53, 169.02, 171.75. HRMS calcd for $C_{20}H_{18}O_6$ 355.1182, found 355.1170.

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References

- (1) (a) Barre´-Sinoussi, F.; Chermann, J. C.; Rey, F.; Nugeyre, M. T., Chamaret, S.; Gruest, J.; Dauguet, C.; Axler-Blin, C.; Vézinet-Brun, F.; Rouzioux, C.; Rozenbaum, W.; Montagnier, L. Isolation of a T-lymphotropic retrovirus from a patient at risk for acquired immune deficiency syndrome. *Science* **1983**, 220 , 868–871. (b) immune deficiency syndrome. *Science* **¹⁹⁸³**, *²²⁰*, 868-871. (b) Gallo, R. C.; Sarin, P. S.; Gelman, E. P.; Robert-Guroff, M.; Richardson, E.; Kalyanaraman, V. S.; Mann, D.; Sidhu, G. D.; Stahl, R. E.; Zolla-Pazner, S.; Leibowitch, J.; Popovic, M. Isolation of human T-cell leukemia virus in acquired immune deficiency syndrome (AIDS). *Science* **¹⁹⁸³**, *²²⁰*, 865-867. (c) Clavel, F.; Guétard, D.; Brun-Vézinet, F.; Chamaret, S.; Rey, M.-A.; Santos-Ferreira, M. O.; Laurent, A. G.; Dauguet, C.; Katlama, C.; Rouzioux, C.; Klatzmann, D.; Champalimaud, J. L.; Montagnier, L. Isolation of a new human retrovirus from west
Africa patients with AIDS. Science 1986, 233, 343-346.
- Africa patients with AIDS. *Science* **¹⁹⁸⁶**, *²³³*, 343-346. (2) (a) McQuade, T. J.; Tomaselli, A. G.; Liu, L.; Karacostas, V.; Moss, B.; Sawyer, T. K.; Heinrikson, R. L.; Tarpley, W. G. A synthetic HIV-1 protease inhibitor with antiviral activity arrests HIV-like particle maturation. *Science* **¹⁹⁹⁰**, *²⁴⁷*, 454-456. (b) Seelmeier, S.; Schmidt, H.; Turk, V.; von der Helm, K. Human immunodeficiency virus has an aspartic-type protease that can be inhibited by pepstatin A. *Proc. Natl. Acad. Sci. U.S.A*. **1988**,
- *⁸⁵*, 6612-6616. (3) (a) Kohl, N. E.; Emini, E. A.; Schleif, W. A.; Davis, L. J.; Heimbach, J. C.; Dixon, R. A. F.; Scolnick, E. M.; Sigal, I. S. Active human immunodeficiency virus protease is required for viral infectivity. *Proc. Natl. Acad. Sci. U.S.A*. **¹⁹⁸⁸**, *⁸⁵*, 4686- 4690. (b) Gottlinger, H. G.; Sodroski, J. G.; Haseltine, W. A. Role of capsid precursor processing and myristoylation in morphogenesis and infectivity of human immunodeficiency virus type 1. Proc. Natl. Acad. Sci. U.S.A. 1989, 86, 5781-5785. (c) Peng, 1. *Proc. Natl. Acad. Sci*. *U.S.A*. **¹⁹⁸⁹**, *⁸⁶*, 5781-5785. (c) Peng, C.; Ho, B. K.; Chang, T. W.; Chang, N. T. Role of human immunodeficiency virus type 1-specific protease in core protein maturation and viral infectivity. *J. Virol*. **¹⁹⁸⁹**, *⁶³*, 2550-2556.
- (4) Sham, H. L.; Chen, X. HIV protease inhibitors in early development. *Exp. Opin. Invest. Drugs* **¹⁹⁹⁶**, *⁵*, 977-983.
- (5) (a) Naesens, L.; De Clercq, E. Present status of HIV protease inhibitors in the control of HIV infection. *Exp. Opin. Invest. Drugs* **1996**, *5*, 153. (b) De Clercq, E. Toward improved anti-HIV chemotherapy: Therapeutic strategies for intervention with HIV infections. *J. Med. Chem*. **¹⁹⁹⁵**, *³⁸*, 2491-2517. (c) Red-

shaw, S. Inhibitors of HIV proteinase. *Exp. Opin. Invest. Drugs* **¹⁹⁹⁴**, *³*, 273-286. (d) Wlodawer, A.; Erickson, J. Structure-based inhibitors of HIV-1 protease. W. *Annu. Rev. Biochem*. **1993**, *62*, ⁵⁴³-585. (e) Meek, T. D. Inhibitors of HIV-1 protease. *J. Enzyme Inhib*. **¹⁹⁹²**, *⁶*, 65-98. (f) Norbeck, D. W.; Kempf, D. HIV protease inhibitors. *J. Annu. Rep. Med. Chem*. **¹⁹⁹¹**, *²⁶*, 141- 150. (g) Huff, J. R. HIV Protease: A novel chemotherapeutic target for AIDS. *J. Med. Chem*. **¹⁹⁹¹**, *³⁴*, 2305-2314. (h) Kempf, D. J.; Sham, H. L. HIV Protease inhibitors. *Curr. Pharm. Design* **¹⁹⁹⁶**, *²*, 225-246.

- (6) Greer, J.; Erickson, J. W.; Baldwin, J. J.; Varney, M. D. Application of the three-dimensional structures of protein target molecules in structure-based drug design. *J. Med. Chem*. **1994**, *³⁷*, 1035-1054.
- (7) Niklasson, G.; Kvarnström, J.; Classon, B.; Nillroth, U.; Danielson, U. H.; Karlén, A.; Hallberg, A.; Samuelsson, B. Synthesis of *C*² symmetric potential inhibitors of HIV-1 protease from D-mannitol. *J. Carbohydr. Chem*. **¹⁹⁹⁶**, *¹⁵*, 555-569.
- Zuccarello, G.; Bouzide, A.; Kvarnström, I.; Niklasson, G.; Svensson, S. C. T.; Brisander, M.; Classon, B.; Danielson, U. H.; Nillroth, U.; Karlén, A.; Hallberg, A.; Samuelsson, B. HIV-1 Protease Inhibitors Based on Acyclic Carbohydrates. *J. Org. Chem.* **¹⁹⁹⁸**, *⁶³*, 4898-4906.
- (9) (a) Kim, C. U.; McGee, L. R.; Krawczyk, S. H.; Harwood: E.; Harada, Y.; Swaminathan, S.; Bischofberger, N.; Chen, M. S.; Cherrington, J. M.; Xiong, S. F.; Griffin, L.; Cundy, K. C.; Lee, A.; Yu, B.; Gulnik, S.; Erickson, J. W. New series of potent, orally bioavailable, nonpeptidic cyclic sulfones as HIV-1 protease inhibitors. *J. Med. Chem.* **1996**, 39, 3431-3434. (b) Hultén, J.; Bonham, N. M.; Nillroth, U.; Hansson, T.; Zuccarello, G.; Bouzide, A.; Åqvist, J.; Classon, B.; Danielson, U. H.; Karlén, A.; Kvarnström, I.; Samuelsson, B.; Hallberg, A. Cyclic HIV-1 protease inhibitors derived from mannitol: Synthesis, inhibitory potencies, and computational predictions of binding affinities. *J. Med. Chem.* **1997**, *40*, 885–897. (c) Qian, X.; Morís-Varas, F.;
Fitzgerald, M. C.; Wong, C. H. C2-symmetrical tetrahydroxyazepanes as inhibitors of glycosidases and HIV/FIV proteases. *Bioorg. Med. Chem*. **¹⁹⁹⁶**, *⁴*, 2055-2069.
- (10) Kempf, D. J. Design of symmetry-based peptidomimetic inhibitors of human immunodeficiency virus protease. *Methods Enzymol.* **¹⁹⁹⁴**, *²⁴¹*, 334-354. (11) Lam, P. Y.; Jadhav, P. K.; Eyermann, C. J.; Hodge, C. N.; Ru,
- Y.; Bacheler, L. T.; Meek, J. L.; Otto, M. J.; Rayner, M. M.; Wong, Y. N.; Chang, C.-H.; Weber, P. C.; Jackson, D. A.; Sharpe, T. R.; Erickson-Viitanen, S. Rational design of potent, bioavailable, nonpeptide cyclic ureas as HIV protease inhibitors. *Science* **1994**,
- *²⁶³*, 380-384. (12) SYBYL ver. 6.1, Tripos Associates, England.
- (13) (a) Erickson, J.; Neidhart, D. J.; VanDrie, J.; Kempf, D. J.; Wang, X. C.; Norbeck, D. W.; Plattner, J. J.; Rittenbouse, J. W.; Turon, M.; Wideburg, N.; Kohlbrenner, W. E.; Simmer, R.; Helfrich, R.; Paul, D. A.; Knigge, M. Design, activity, and 2.8.ANG. crystal structure of a C2 symmetric inhibitor complexed to HIV-1 protease. *Science* **¹⁹⁹⁰**, *²⁴⁹*, 527-533. (b) Bernstein, F. C.; Koetzle, T. F.; Williams, G. J. B.; Meyer, Jr., E. F.; Brice, M. D.; Rodgers, J. R.; Kennard, O.; Shimanouchi, T.; Tasumi, M. The Protein Data Bank: a computer-based archival file for macromolecular structures. *J. Mol. Biol*. **¹⁹⁷⁷**, *¹¹²*, 535-542. (c) Abola, E.; Bernstein, F. C.; Bryant, S. H.; Koetzle, T. F.; Weng, J. Protein Data Bank. In *Crystallographic Databases* - *Information Content, Software Systems, Scientific Applications*; Allen, F. H., Bergerhoff, G., Sievers, R., Eds.; Data Commission of the International Union of Crystallography: Bonn, 1987; pp 107- 132.
- (14) Le Merrer, Y.; Duréault, A.; Greck, C.; Micas-Languin, D.; Gravier, C.; Depezay, J. C. Synthesis of diepoxides and diaziridines, precursors of enantiomerically pure alpha-hydroxy and alpha-amino aldehydes or acids, from D-mannitol. *Heteroc*. **1987**, *²⁵*, 541-548.
- (15) Chaudhary, S. K.; Hernandez, O. 4-Dimethylaminopyridine: an efficient and selective catalyst for the silylation of alcohols. *Tetrahedron Lett*. **¹⁹⁷⁹**, 99-102.
- (16) Bowers, A.; Halsall, T. G.; Jones, E. R. H.; Lemin, A. J. The chemistry of the triterpenes and related compounds. Part XVIII. Elucidation of the structure of polyporenic acid C. *J. Chem. Soc*. **1953**, 2548.
- (17) Andersson, F.; Samuelsson, B. Pyridinium dichromate-acetic acid anhydride: A new and highly efficient reagent for the oxidation of alcohols. *Carbohydr. Res*. **¹⁹⁸⁴**, *¹²⁹*, C1-C3.
- (18) (a) Anelli, P. L.; Biffi, C.; Montanari, F.; Quici, S. Fast and selective oxidation of primary alcohols to aldehydes or to carboxylic acids and of secondary alcohols to ketones mediated by oxoammonium salts under two-phase conditions. *J. Org. Chem*. **¹⁹⁸⁷**, *⁵²*, 2559-2562. (b) Anelli, P. L.; Banfi, S.; Montanari, F.; Quici, S. Oxidation of diols with alkali hypochlorites catalyzed by oxammonium salts under two-phase conditions. *J. Org. Chem*. **¹⁹⁸⁹**, *⁵⁴*, 2970-2972.
- (19) Poirier, V.; Calleman, C.-J. The nucleophilic reactivity of valine methylamide. A model of the N-terminal valine residues of hemoglobin Acta Chem. Scand. Ser. B 1983, 37, 817–822.
- hemoglobin *Acta Chem. Scand. Ser. B* **1983**, 37, 817–822.
(20) Ogura, H.; Kobayashi, T.; Shimizu, K.; Kawabe, K.; Takeda, K.
A novel active ester synthesis reagent (N,N'-disuccinimidyl carbonate). *Tetrahedron Lett.* **¹⁹⁷⁹**, 4745-4746.
- (21) (a) Knorr, R.; Trzeciak, A.; Bannwarth, W.; Gillessen, D. New coupling reagents in peptide chemistry. *Tetrahedron Lett.* **1989**, *³⁰*, 1927-1930. (b) Bannwarth, W.; Schmidt, D.; Stallard, R. L.; Hornung, C.; Knorr, R.; Müller, F. Bathophenanthroline-ruthenium(II) complexes as nonradioactive labels for oligonucleotides which can be measured by time-resolved fluorescence techniques.
Helv. Chim. Acta **1988**, 71, 2085–2099. (c) Bannwarth, W.;
Knorr. R. Formation of carboxamides with NNN'N'-tetra-Knorr, R. Formation of carboxamides with N,N,N′,N′-tetramethyl(succinimido)uronium tetrafluoroborate in aqueous organic solvent systems. *Tetrahedron Lett.* **¹⁹⁹¹**, *³²*, 1157-1160.
- (22) (a) Marsham, P. R.; Jackman, A. L.; Barker, A. J.; Boyle, F. T.; Pegg, S. J.; Wardleworth, J. M.; Kimbell, R.; O'Connor, B. M.; Calvert, A. H.; Hughes, L. R. Quinazoline antifolate thymidylate synthase inhibitors: Replacement of glutamic acid in the C2 methyl series. *J. Med. Chem*. **¹⁹⁹⁵**, *³⁸*, 994-1004. (b) Kisfaludy, L.; Schón, I.; Szirtes, T.; Nyéki, O.; Lów, M. Novel and rapid
- peptide synthesis. *Tetrahedron Lett.* **1974**, 1785-1786.
(23) Fernandez, J. M. G.; Mellet, C. O.; Marín, A. M.; Fuentes, J. Mild and efficient procedure to remove acetal and dithioacetal protecting groups in carbohydrate derivatives using 2,3-dichloro-5,6-dicyano-1,4-benzoquinone. *Carbohydr. Res.* **¹⁹⁹⁵**, 263-268.
- (24) Linstead, R. P.; Owen, L. N.; Webb, R. F. Elimination reactions of esters. Part III. Derivatives of dibasic polyhydroxy-acids. *J. Chem. Soc.* **¹⁹⁵³**, 1225-1231.
- (25) Iversen, T.; Bundle, D. R. Benzyl trichloroacetimidate, a versatile reagent for acid-catalyzed benzylation of hydroxy-groups. *J. Chem. Soc., Chem. Commun.* **¹⁹⁸¹**, 1240-1241.
- (26) Markgren, P.-O.; Lindgren, M.; Nillroth, U.; Danielson, U. H. Identification of a reversible high affinity interaction between $Cu2+$ and HIV-1 proteinase residue His- 69 resulting in allosteric inhibition. Submitted for publication.
- (27) Nillroth, U.; Besidsky, Y.; Classon, B.; Chattopadhyaya, J.; Ugi, I.; Danielsson, U. H. Specific interaction between HIV-1 proteinase and 5′-phosphate peptidomimetic derivatives of nucleoside analogues. *Drug Des. Discovery* **1995**, *13*, 43.
- (28) Nillroth, U.; Vrang, L.; Markgren, P.-O.; Hultén, J.; Hallberg, A.; Danielson, U. H. Human immunodeficiency virus type 1 proteinase resistance to symmetric cyclic urea inhibitor analogues. *Antimicrob. Agents Chemother.* **¹⁹⁹⁷**, *⁴¹* (11), 2383-2388.
- (29) Classon, B.; Kvarnström, I.; Samuelsson, B.; Unge, T. Unpublished results.
- (30) Weislow, O. S.; Kiser, R.; Fine, D. L.; Bader, J.; Shoemaker, R. H.; Boyd, M. R. New soluble-formazan assay for HIV-1 cytopathic effects: Application to high-flux screening of synthetic and natural products for AIDS-antiviral activity. *J. Natl. Cancer Inst*.
- **¹⁹⁸⁹**, *⁸¹*, 577-586. (31) Santi, D. V.; Danenberg, P. V. Phenylalanyl transfer ribonucleic acid synthetase from *Escherichia coli*. Analysis of the phenyl-
- alanine binding site. *Biochemistry* **¹⁹⁷¹**, *¹⁰*, 4813-4820. (32) Plucinski, T.; Rolka, K.; Baran, L.; Przegalinski, E.; Kupryszewski, G. Opiate-like peptides. Part III. New amides of Metenkephalin and [D-Ala]2-Met-enkephalin. *Polish J. Chem*. **1983**, *57*, 887.
- (33) Pavlik, M.; Kluh, I.; Pavlikova, F.; Vasickova, S.; Kostka, V. Preparation, spectral and physicochemical characteristics of methylamide N^{α} -phenylthiocarbamoyl derivatives of naturally occurring amino acids. *Collect. Czech. Chem. Commun.* **1989**, *⁵⁴*, 1940-1954.

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